

ASYMMETRIC HYDROLYSIS OF *dl* 3-(1-BENZOYLOXYETHYL)-AZETIDINONE
DERIVATIVE AND CHEMICAL CONVERSION TO CARBAPENEM

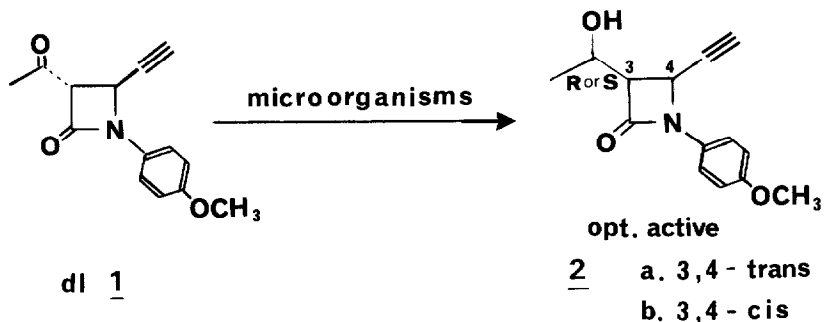
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Summary: Asymmetric hydrolysis of *dl* R* benzoate (*dl* 3) was achieved effectively by *Bacillus subtilis* to give the optically-active R-hydroxyethyl azetidinone 8, and this product 8 was converted chemically into the known β -azetidinyl acetyl thioester derivative 12, which was further converted into carbapenem derivative 14.

In a previous communication¹⁾ we reported the microbial transformation of *dl* 3-acetylazetidinone derivative 1, and obtained optically active R- or S-hydroxyethyl derivative 2 of 3,4-*trans* or -*cis* configuration, depending on the microorganisms employed. In the microorganisms so far examined, only *Saccharomyces rosei* was effective to produce the desired 3S,4S-(R)-hydroxyethyl azetidinone (2a), but the optical yield was disappointingly low. (Scheme 1).

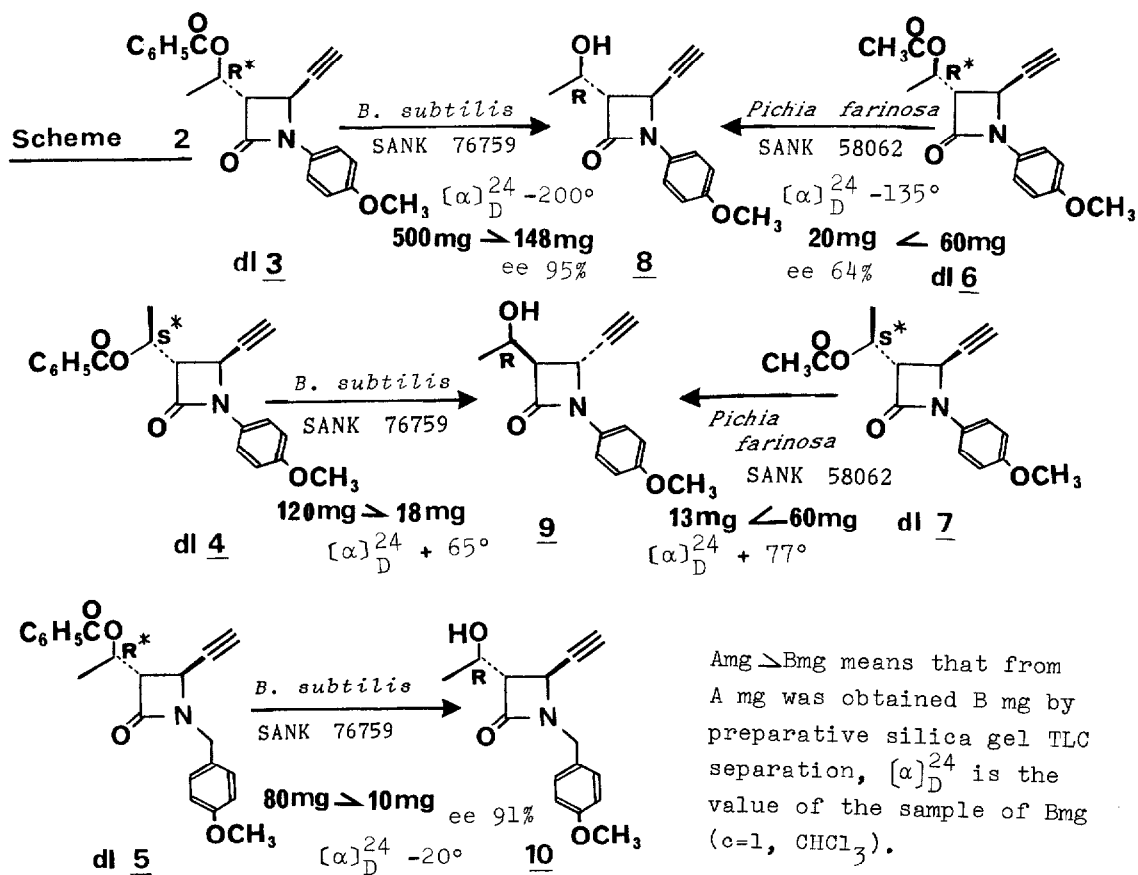
Scheme 1



Here we wish to report the microbial asymmetric hydrolysis of esters in *dl* monocyclic β -lactams. The procedure is as follows: after feeding 500 mg of *dl* R*-benzoate (*dl* 3)²⁾ in 1 l of culture medium of *Bacillus subtilis* at an exponential phase, followed by incubation at 37°C for 24 hr, the products were extracted with ethylacetate. Evaporation of the solvent gave the crude products (520 mg), which were separated on silica gel preparative TLC(2mm), c-Hex.: EtOAc = 1:1, to afford 148 mg of the optically-

active R-hydroxyethyl derivative **8** (mp 133°C, $[\alpha]_D^{24} -200^\circ$ (c=1, CHCl₃), ee 95%. For S*-benzoate (*dl* **4**) the product was R-hydroxyethyl derivative **9**, but the optical and conversion yields were low.

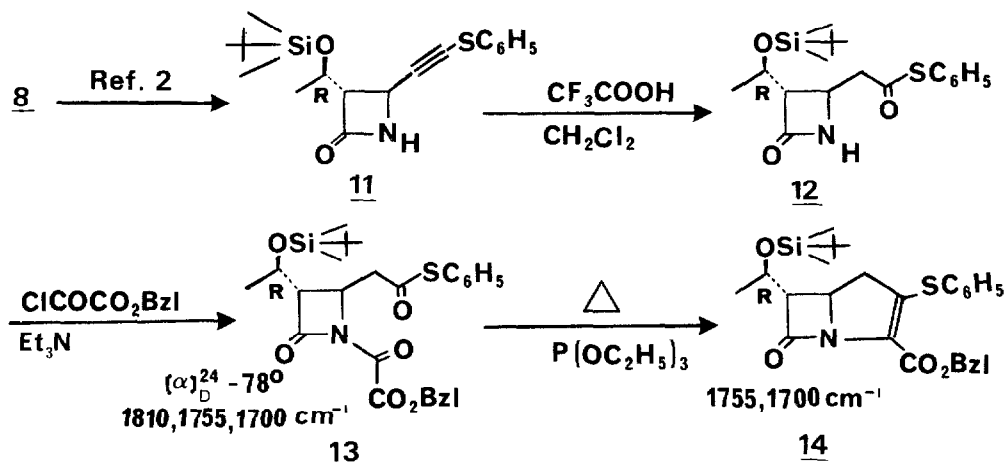
For acetates (*dl* **6** and *dl* **7**) *Pichia farinosa* was more effective than *Bacillus subtilis*, but the optical yields were not satisfactory. (Scheme 2). N-*p*-anisyl- R* - Benzoate (*dl* **5**) gave R-hydroxyethyl derivative **10**, $[\alpha]_D^{24} -20^\circ$, ee 91% (cf. standard **10** derived from 6-APA, $[\alpha]_D^{24} -22^\circ$), Rf=0.22 (c-Hex:AcOEt=1:1). NMR(CDCl₃, 60 MHz) δ : 1.24(3H, d, J=6Hz), 2.39(1H, d, J=2Hz), 3.22(1H, dd, J=5 and 2Hz), 3.70(3H, s), 3.9-4.4(1H, m), 3.97(1H, d, J=15Hz), 4.59(1H, d, J=15Hz), 6.7-7.3(4H, A₂B₂).²⁾ It is noteworthy to point out here that in both *Bacillus subtilis* and *Pichia farinosa*, all of the hydrolyzed products have a hydroxyethyl group of R-configuration. (Scheme 2).



Pig liver esterase (PLEase), which is very commonly used for the asymmetric hydrolysis of C₂-symmetric diesters³⁾, seemed to be very attractive to our β -lactam esters, but from 60 mg of R*-benzoate (*dl* **3**) only 12.4 mg of the hydrolyzed product **8**, ($[\alpha]_D^{24} -86^\circ$) was obtained, which means PLEase was not effective in this case.

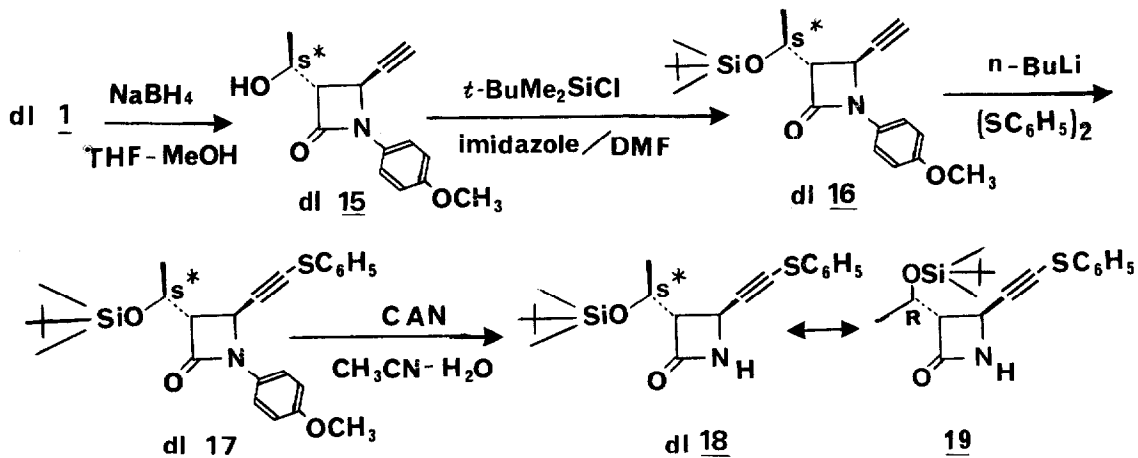
As described above the best result so far obtained was from the application of the R*-benzoate (*dl* 3) to *Bacillus subtilis*, and we further transformed this optically-active R-hydroxyethyl derivative 8 into the carbapenem derivative 14 as shown in scheme 3. The R-hydroxyethyl derivative 8 hydrolyzed by *Bacillus subtilis* was transformed into phenylthio derivative 11 (mp 78°C, $[\alpha]_D^{24} +46^\circ$) by the same procedure described in Ref. 2) in 42 % yield from 8. Hydration of the triple bond was smoothly achieved by treatment with 5 eq. of trifluoroacetic acid to afford the known β -azetidinyl-acetylthio-ester derivative 12⁴⁾ From this compound 12 by the reported procedure (benzyl oxalyl chloride/Et₃N, and then heating in xylene in the presence of triethyl phosphite) the carbapenem derivative 14 was obtained without any difficulties⁵⁾. (Scheme 3).

Scheme 3



Reference and Footnotes

- 1) K. Hirai and A. Naito, *Tetrahedron Lett.*, 1989 in press.
- 2) The preparation of starting esters and their structure determination were performed as follows: Sodium borohydride reduction of *dl* 3-acetylazetidinone (1)¹⁾ in THF and methanol gave the S*-alcohol 15 (mp 109°C) and R*-alcohol in a ratio of 3:2. The structure of the easily crystallizable S*-alcohol 15 was confirmed unequivocally by the comparison of its chemically converted derivative (*dl* 18, mp 93°C) with the standard compound 19 prepared from 6-APA by the reported procedure⁴⁾. *dl* 15: NMR (60 MHz, CDCl₃) δ : 1.37(3H, d, J=6Hz), 2.55(1H, d, J=2Hz), 2.67(1H, OH), 3.40(1H, dd, J=4 and 2Hz), 3.75(3H, s), 3.9-4.4(1H, m), 4.45(1H, t, J=2Hz), 6.75-7.60(4H, A₂B₂). IR (Nujol) cm^{-1} : 3550, 3220, 2130, 1732, 1520. *dl* 16: 0.1(6H, s), 0.8(9H, s), 1.33(3H, d, J=6Hz), 2.46(1H, d, J=2Hz), 3.42(1H, 1dd, J=3 and 4.5Hz), 3.75(3H, s), 4.0-4.4(1H), 4.38(1H, t, J=2Hz), 6.6-7.5(4H, A₂B₂). *dl* 17: 0.08(6H, s), 0.81(9H, s), 1.38(3H, d, J=6Hz), 3.48(1H, dd, J=3 and 4Hz), 3.74(3H, s), 4.0-4.4(1H), 4.62(1H, d, J=3Hz), 7.28(5H, br s), 6.65-7.60(4H, A₂B₂). *dl* 18: 0.08(6H, s), 0.88(9H, s), 1.33(3H, d, J=6Hz), 3.50(1H, br t, J=3Hz), 4.1-4.40(1H, m), 4.36(1H, d, J=3Hz), 6.1(1H, br s), 7.25-7.55(5H). Mp 93°C.



The starting esters (**dl 3**-**dl 7**): the S^* -alcohol (**dl 15**) was benzoylated ($\text{C}_6\text{H}_5\text{COCl}/\text{Et}_3\text{N}$) to **dl 4** and was also converted into the R^* -benzoate (**dl 3**), mp 101°C by Mitsunobu reaction ($\text{C}_6\text{H}_5\text{CO}_2\text{H}, (= \text{NCO}_2\text{Et})_2, \text{P}(\text{C}_6\text{H}_5)_3$). In the case of N -*p*-anisyl derivative the R^* -benzoate (**dl 5**), M^+ 363 was prepared by the same reaction sequence for the preparation of **dl R-benzoate **3**. The acetates (**dl 6** and **dl 7**) were prepared analogously.**

dl 3: mp 101°C , Rf=0.5(CH_2Cl_2). IR (Nujol) cm^{-1} : 3280, 2140, 1745, 1720, 1608, 1590. NMR(60 MHz, CDCl_3) δ : 1.55(3H, d, J=6.5Hz), 2.55(1H, d, J=2.5Hz), 3.60(1H, dd, J=2.5 and 6.5Hz), 3.70(3H, s), 4.60(1H, t, J=2.5Hz), 5.46(1H, q, J=6.5Hz), 6.7-7.6(7H), 7.8-8.0(2H). **dl 4**: Rf=0.61(CH_2Cl_2), NMR: 1.59(3H, d, J=6.5Hz), 2.55(1H, d, J=2.5Hz), 3.70(3H, s), 4.38(1H, t, J=2.5Hz), 3.6-3.73(1H), 5.53(1H, dq, J=6.5 and 3Hz), 6.74-7.9(9H). **dl 5**: Rf=0.25(c-Hex:AcOEt=2:1), NMR: 1.43(3H, d, J=6Hz), 2.51(1H, d, J=2Hz), 3.49(1H, dd, J=6 and 2Hz), 3.73(3H, s), 4.0(1H, d, J=15Hz) and 4.70(1H, d, J=15Hz), 5.40(1H, q, J=6 Hz), 6.6-6.7(7H), 7.6-7.9(2H). **dl 6**: Rf=0.26(CH_2Cl_2), NMR δ : 1.40(1H, d, J=6.5Hz), 2.00(3H, s), 2.55(1H, d, J=2Hz), 3.45(1H, dd, J=6.5 and 2Hz), 3.76(3H, s), 4.50(1H, t, J=2Hz), 5.27(1H, q, J=6.5Hz), 6.7-7.6(4H, A_2B_2). **dl 7**: Rf=0.34(CH_2Cl_2), NMR δ : 1.42(3H, d, J=6.5Hz), 2.0(3H, s), 2.55(1H, d, J=2Hz), 3.76(3H, s), 3.57(1H, dd, J=5 and 2.5Hz), 4.31(1H, t, J=2.5Hz), 5.30(1H, dq, J=6.5 and 5Hz), 6.7-7.6(4H, A_2B_2). (-)**8**: NMR δ : 1.27(3H, d, J=6Hz), 2.55(1H, d, J=2Hz), 3.38(1H, dd, J=4 and 2Hz), 3.75(3H, s), 4.1-4.5(1H), 4.60(1H, t, J=2Hz), 6.7-7.6(4H, A_2B_2).

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(Received in Japan 15 October 1988; accepted 27 February 1989)